#### AerE545 / AerE445 class notes

**LECTURE 25:** 

**Molecular Tagging Techniques** 

**Part - 02** 

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# $\square$ PLIF - intensity at point $(x_0, y_0)$

$$H_f(x_0, y_0) = I(x_0, y_0) A\Phi(T) \varepsilon(T) C(x_0, y_0)$$

camera

 $H_f(x_0, y_0)$ : measured fluorescence intensity

A: the fraction of the fluorescence light collected by camera.

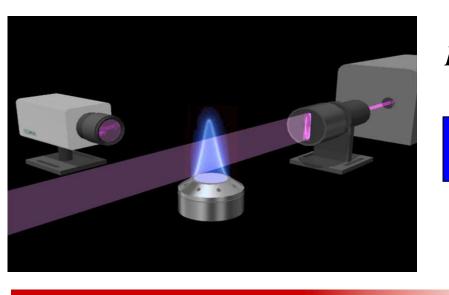
Φ: quantum efficiency,

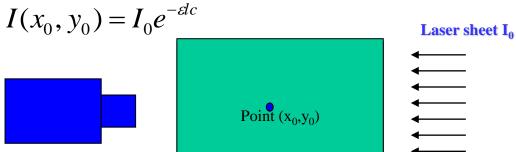
L: the length of the sampling volume along the path of excitation beam

**E** molar absorption coefficient

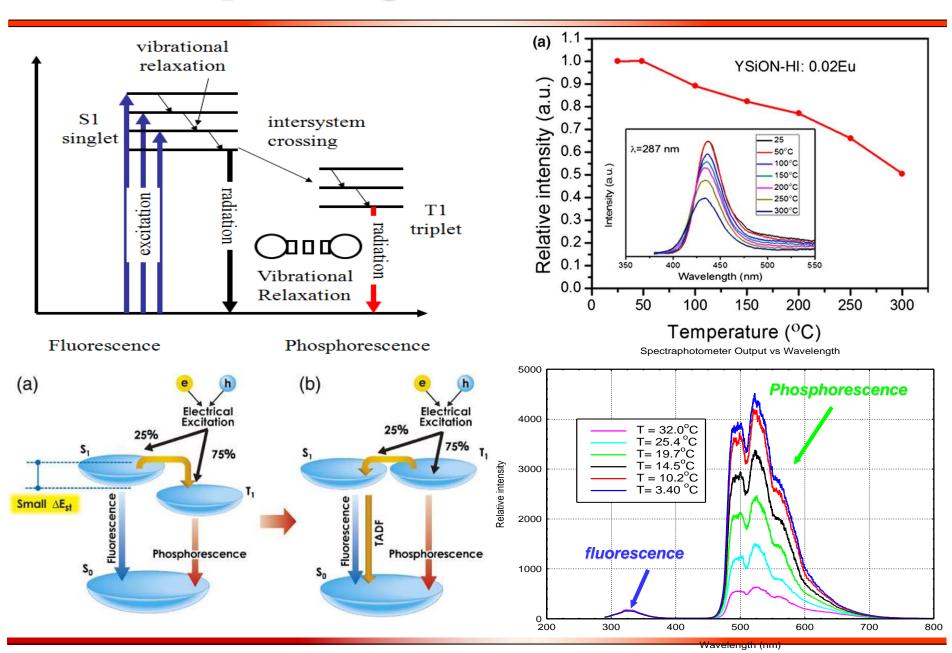
 $C(x_0,y_0)$ : the molar concentration of the fluorescent dye.

 $I(x_0,y_0)$ : the intensity of excitation light beam at the point  $(x_0,y_0)$ 

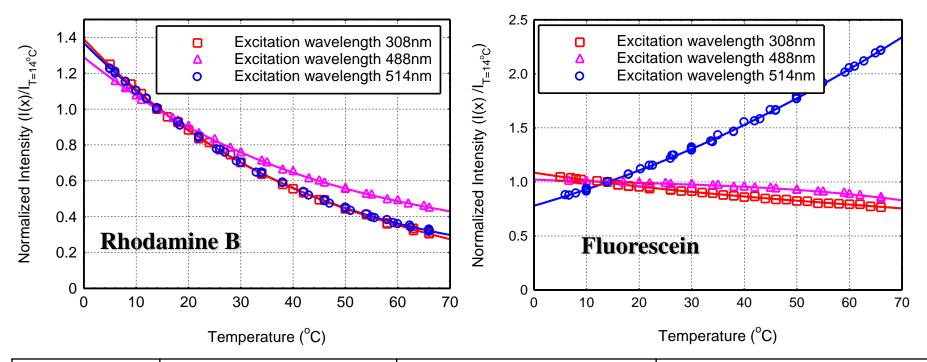




## ☐ Thermal Quenching of Photoluminescence

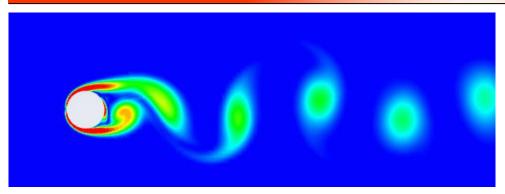


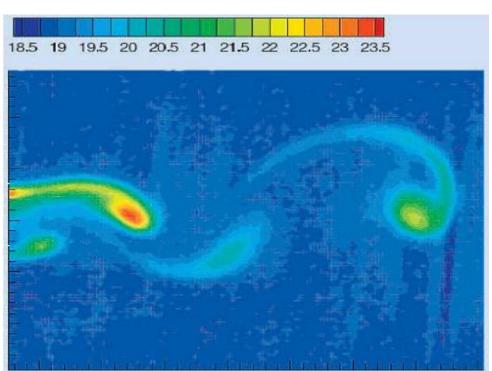
# LIF WITH TEMPERATURE DEPENDENT FLUORESCENT DYES: RHODAMINE B AND FLUORESCEIN

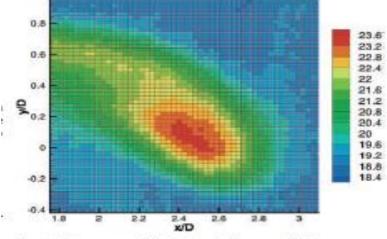


	308nm UV laser	488nm Argon-ion laser	514nm Argon-ion laser
Rhodamine B	-1.55% K <sup>-1</sup> (5°C ~ 65°C)	-1.17% K <sup>-1</sup> (6°C ~ 66°C) *Sakakibara and Adrian -2.3 % K <sup>-1</sup> (15°C ~ 40°C)	-1.48% per K <sup>-1</sup> (6°C ~ 66°C) *Coppeta and Roger -1.54 % K <sup>-1</sup> (20°C ~ 60°C)
Fluorescein	-0.45% K <sup>-1</sup> (5°C ~ 65°C)	-0.25% K <sup>-1</sup> (6°C ~ 66°C) *Coppeta and Roger -0.16 % K <sup>-1</sup> (20°C ~ 60°C)	+2.25% K <sup>-1</sup> (6°C ~ 66°C) *Coppeta and Roger +2.43 % K <sup>-1</sup> (20°C ~ 60°C)

#### QUANTIFICATION OF HEAT TRANSFER AROUND A HEATED CYLINDER







Pig. 10. Temperature distribution in the warm blob

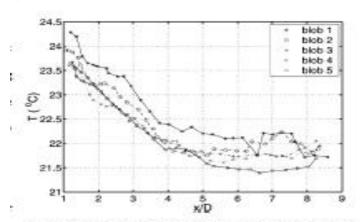


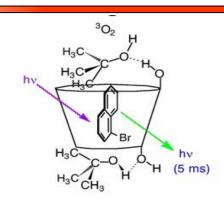
Fig. 11. The maximum temperature in the warm blob as function of the downstream position

• Seuntiëns, H. J.; Kieft, R. N.; Rindt, C. C. M.; van Steenhoven, A. A., "2D temperature measurements in the wake of a heated cylinder using LIF", Experiments in Fluids, Volume 31, Issue 5, pp. 588-595 (2001).

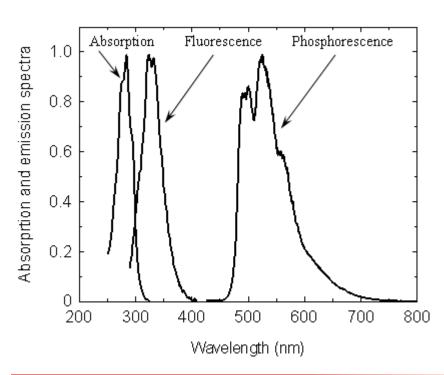
#### ■ Long Lifetime Phosphorescent Molecular Tracers

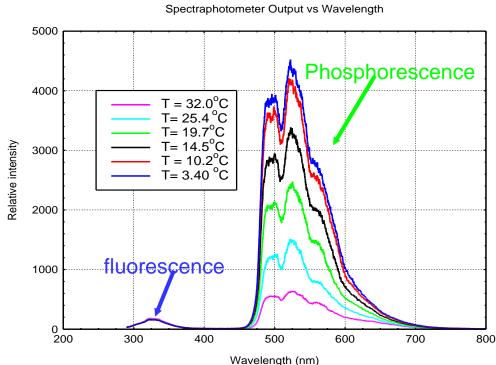
#### Phosphorescent triplex (1-BrNp • $G\beta$ -CD • ROH)

- 1. Bromonaphthalene (1-BrNp)
- 2. Cyclodextrin (G $\beta$ -CD)
- 3. Alcohols (ROH)



# Molecular structure of the phosphorescent triplex





#### Technical Basis for Temperature Measurement

According to quantum theory, the decay of phosphorescence emission intensity
 (I<sub>em</sub>) follows an exponential law:

$$I_{em} = I_o e^{-t/\tau} = I_i C \varepsilon \Phi_p e^{-t/\tau}$$

*I<sub>o</sub>*: Initial phosphorescence intensity:

$$I_o = I_i C \varepsilon \Phi_p$$

*I<sub>i</sub>*: the local incident laser intensity

C: concentration of phosphorescence dye

 $\varepsilon$ : the absorption coefficient

 $\Phi_{p}$ : phosphorescence quantum yield, temperature-dependent

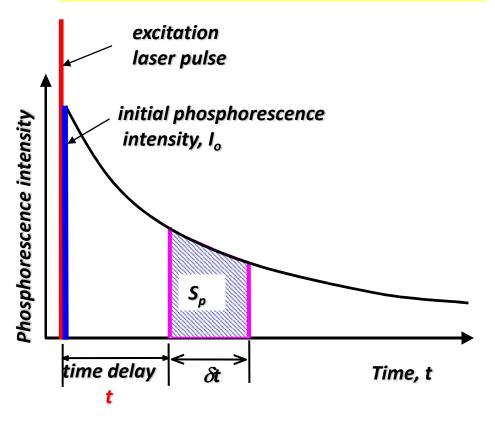
 $\tau$ : phosphorescence lifetime, which refers to the time when the intensity drops to 37% (i.e. 1/e) of the initial intensity ( $I_0$ ), temperature-dependant.

$$\begin{array}{c} \Phi = \Phi(T) \\ \tau = \tau(T) \end{array} \right\} \quad \Rightarrow \quad \begin{cases} I_o = I_o(T) \\ I_{em} = I_{em}(T) \end{cases}$$

#### Technical Basis for Temperature Measurement

Capturing the phosphorescence emission at time t using a gated CCD detector:

$$S_{p} = \int_{t}^{t+\delta t} I_{em} dt = I_{o}(T) \cdot \tau(T) \left(1 - e^{-\frac{\delta t}{\tau(T)}}\right) e^{-\frac{t}{\tau(T)}}$$
• At a prescribed temperature  $T = T_{o}$ 



$$S_{p_o} = I_o(T_o) \cdot \tau(T_o) \begin{pmatrix} -\frac{\delta t}{\tau(T_o)} \\ 1 - e^{-\frac{\delta t}{\tau(T_o)}} \end{pmatrix} e^{-\frac{t}{\tau(T_o)}}$$

Relative intensity:

$$R = \frac{S_p}{S_{p_o}} = \frac{I_o(T) \cdot \tau(T) \begin{pmatrix} -\frac{\delta t}{\tau(T)} \\ 1 - e^{-\frac{\delta t}{\tau(T)}} \end{pmatrix}}{I_o(T_o) \cdot \tau(T_o) \begin{pmatrix} -\frac{\delta t}{\tau(T_o)} \\ 1 - e^{-\frac{\delta t}{\tau(T_o)}} \end{pmatrix}} e^{-\left[\frac{1}{\tau(T)} - \frac{1}{\tau(T_o)}\right] \cdot \mathbf{t}}$$

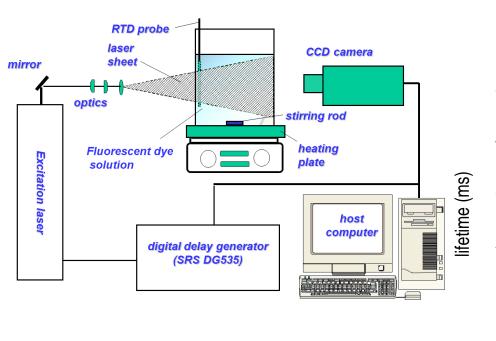
$$= \mathbf{R}(T, \mathbf{t})$$

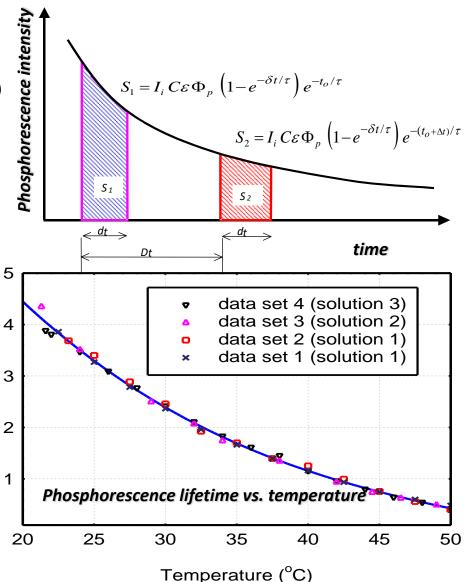
- Relative intensity R is the function of temperature, T, and time delay after laser pulse, t!!!
- The sensitivity for temperature measurement is adjustable by changing the time delay, t!!!

#### □ Phosphorescence Lifetime vs. Temperature

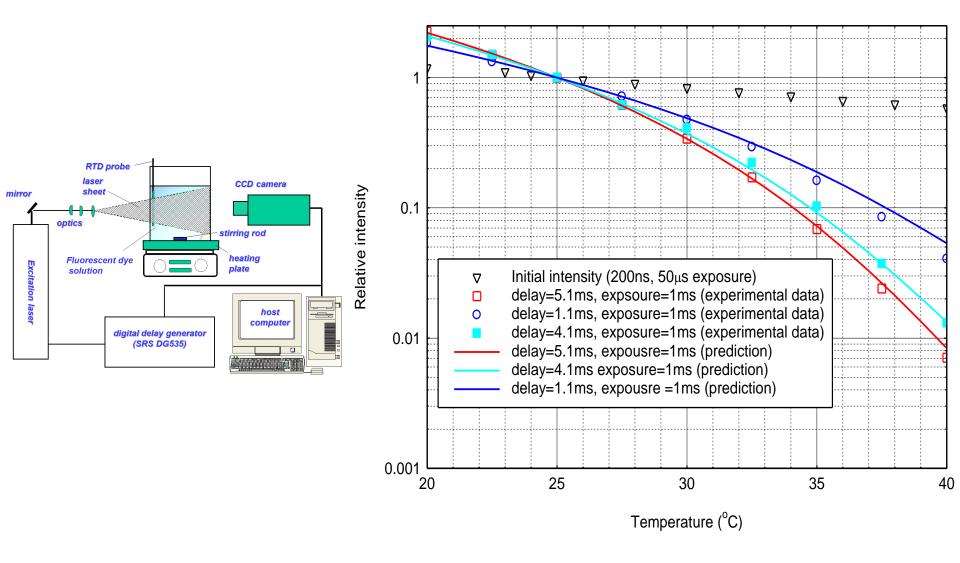
#### • Lifetime imaging technique:

$$\frac{S_2}{S_1} = e^{-\Delta t/\tau} \quad \Rightarrow \quad \tau = \frac{\Delta t}{\ln(S_2/S_1)} \quad \Rightarrow \tau = \tau(T)$$





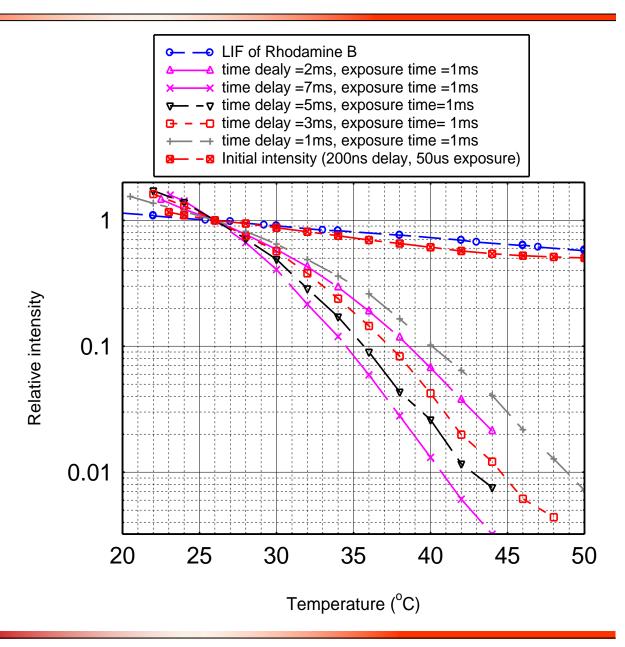
#### ☐ Initial Phosphorescence Intensity vs. Temperature



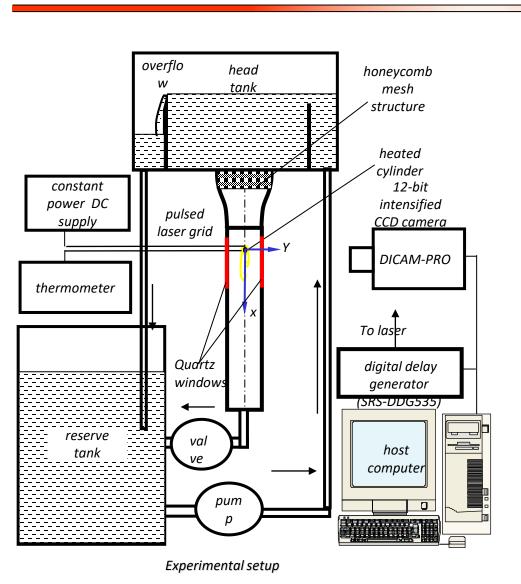
#### □ Calibration Profiles with Different Time Delay

# Sensitivity for temperature measurement at 25 °C

Time delay after laser pulse	Temperature sensitivity (K <sup>-1</sup> )	
Delay=1 ms	5.8 %	
Delay=2 ms	8.9 %	
Delay=3 ms	13.0 %	
Delay=5 ms	15.2 %	
Delay=7 ms	21.5 %	
LIF using Rhodamine B	2.2 %	



#### ■ Demonstration Experimental Setup



Thermocouple **Copper** cylinder Cartridge hadter Thermocouple Mirror Thermal paste Heated cylinder Heated cvlinder Beam Splitter Beam Blocker Quartz Windows Cylindrical Mirror Water Optical setup Rectangular Beam Channe from Excimer UV Laser (308nm)

#### Present experimental parameters:

 $U_{in} = 0.034 \text{ m/s}$ 

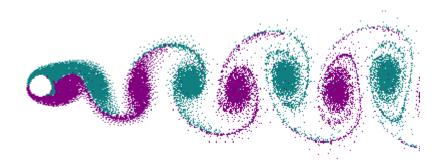
D = 4.76mm

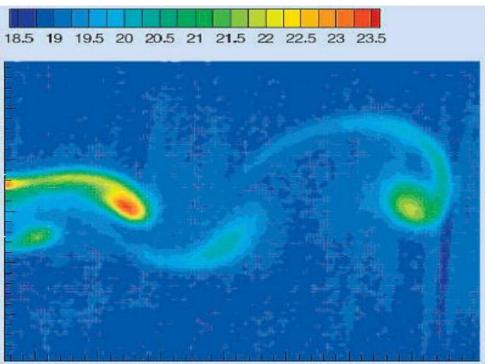
 $T_{fluid} = 24.0 \, \, ^{\circ}\!\! \mathrm{C}$ 

 $T_{cylinder}$ = 57.0 °C

Re = 170

# **QUANTIFICATION OF HEAT TRANSFER AROUND A HEATED CYLINDER**





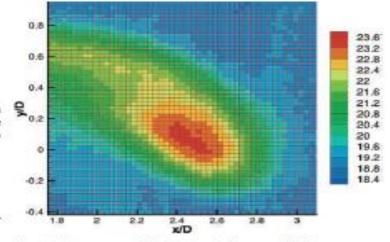


Fig. 10. Temperature distribution in the warm blob

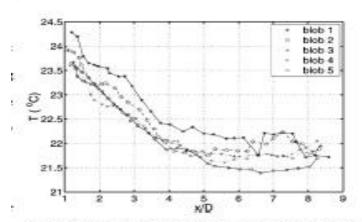
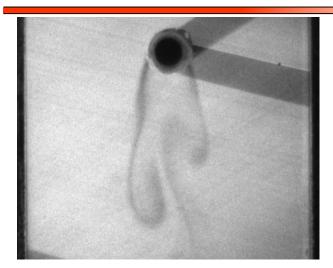


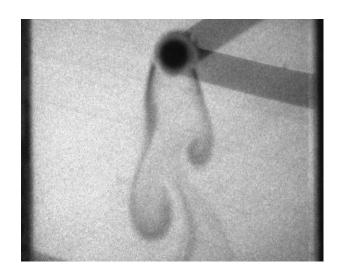
Fig. 11. The maximum temperature in the warm blob as function of the downstream position

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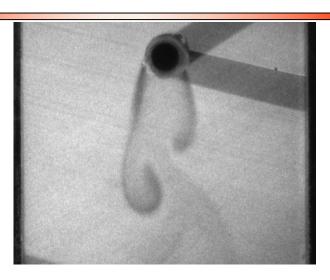
### □ Comparison of Raw Images with Different Time Delay



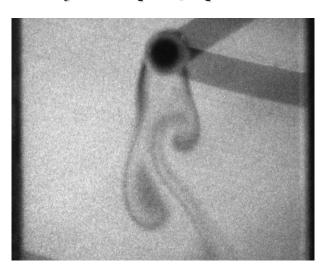
a. 1ms after laser pulse, exposure time 1ms



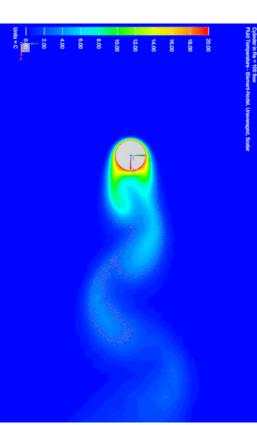
c. 5ms after laser pulse, exposure time 1ms



b. 3ms after laser pulse, exposure time 1ms



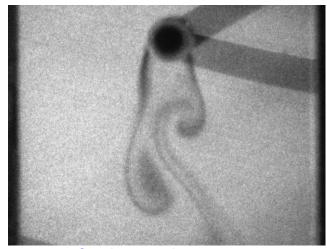
d. 7ms after laser pulse, exposure time 1ms



### ■ Molecular Tagging Thermometry (MTT) results

Relative intensity

$$I_{em} = I_o e^{-t/\tau} = I_i C \varepsilon \Phi_p e^{-t/\tau}$$



a. 7ms after laser pulse, exposure time



b. Background

